

Role of Enamel Matrix Protein Derivatives in Promoting Periodontal Regeneration: Molecular Mechanisms and Biological Insights — A Comprehensive Review

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Abstract

Periodontal regeneration requires the coordinated reformation of cementum, periodontal ligament (PDL), and alveolar bone. Enamel matrix protein derivatives (EMD), primarily composed of amelogenins, have emerged as a biologically active biomimetic agent capable of recapitulating early cementogenesis. This review synthesizes current evidence regarding the biological mechanisms, molecular pathways, cellular

interactions, and regenerative potential of EMD in periodontal therapy.

Historically derived from the enamel matrix secretions of Hertwig’s epithelial root sheath, EMD plays a critical role in cementoblast differentiation and the formation of acellular extrinsic fiber cementum. At the molecular level, EMD activates several signaling cascades, including TGF- β , BMP-2/7, MAPK, and Wnt pathways, which regulate fibroblast proliferation, collagen synthesis, osteogenic differentiation, and extracellular

matrix maturation. EMD also exhibits anti-inflammatory and angiogenic effects by modulating cytokine expression and upregulating VEGF.

Histologic and preclinical studies consistently demonstrate that EMD promotes the formation of new cementum with inserting Sharpey's fibers, functional PDL organization, and alveolar bone formation. Clinical trials further support its effectiveness in intrabony defects, furcation involvement, and soft-tissue procedures. Synergistic outcomes have been reported when EMD is combined with xenografts, collagen matrices, platelet-derived growth factor (PDGF), or biomimetic scaffolds.

While EMD remains a widely accepted biologic for periodontal regeneration, certain limitations—including sensitivity to environmental factors, variability in clinical outcomes, and ethical concerns regarding its porcine origin—necessitate ongoing innovation. Emerging technologies such as recombinant amelogenin peptides, nanostructured carriers, hydrogel-based delivery systems, and tissue-engineered constructs offer promising opportunities to enhance the predictability and versatility of EMD.

In conclusion, EMD serves as a foundational biologic material capable of initiating periodontal regeneration through highly conserved developmental pathways. Its established clinical safety, biologic efficacy, and compatibility with adjunctive regenerative materials support its continued use in modern periodontal therapy, while future advances may further refine its regenerative potential.

Keywords: Enamel Matrix Derivative Amelogenin, Periodontal Regeneration, Cementogenesis, Biomimetics, Extracellular Matrix Proteins, Hertwig's Epithelial Root Sheath, Tissue Engineering, Growth Factors.

Introduction

Periodontal diseases are chronic inflammatory conditions characterized by progressive destruction of the gingiva, periodontal ligament (PDL), cementum, and alveolar bone, ultimately compromising tooth support and oral function. Achieving true periodontal regeneration requires the coordinated reformation of new acellular extrinsic fiber cementum, functional PDL fibers, and supporting alveolar bone—features that clearly differentiate regeneration from reparative healing, which typically results in long junctional epithelium. Traditional regenerative approaches such as guided tissue regeneration (GTR), bone grafting, and barrier membranes have shown variable success, often influenced by defect morphology, surgical technique, and patient-related factors.¹⁻⁴

The limitations of conventional therapies have prompted a shift toward biologically based regenerative materials that can mimic natural developmental processes. Enamel matrix derivative (EMD), commercially available as Emdogain®, is one of the most extensively studied biomimetic agents in periodontal regeneration. Derived from porcine tooth buds and composed predominantly of amelogenins, EMD is designed to emulate epithelial-mesenchymal interactions mediated by Hertwig's epithelial root sheath during root development.^[5-9] Through its effects on cellular adhesion, proliferation, differentiation, and extracellular matrix formation, EMD supports the biological cascade necessary for new cementum, PDL, and alveolar bone formation.

A strong body of histologic, preclinical, and clinical evidence demonstrates that EMD promotes the formation of acellular extrinsic fiber cementum with inserting Sharpey's fibers, organized PDL, and alveolar bone. These regenerative outcomes have been validated in intrabony defects, recession coverage procedures, and

furcation involvement.^[10-14] However, variability exists in clinical response due to factors such as defect anatomy, local wound stability, host healing capacity, and environmental sensitivity of amelogenins.

Given the substantial role of EMD in modern regenerative periodontology, this review provides a comprehensive synthesis of its molecular mechanisms, cellular effects, histologic evidence, and clinical applications. It also highlights emerging biomimetic and engineered approaches aimed at enhancing the predictability and biological performance of EMD.

The primary objective of this review is to present an updated and evidence-based overview of enamel matrix protein derivatives, emphasizing their biologic foundation and future potential in periodontal regeneration.

Historical Background

The biological basis for the use of enamel matrix protein derivatives (EMD) in periodontal regeneration originates from classic developmental studies of tooth root formation. Hertwig's epithelial root sheath (HERS) was identified as the key epithelial structure responsible for guiding root morphogenesis and initiating cementogenesis. Early investigations demonstrated that HERS secretes enamel matrix proteins—including amelogenin, enamelin, and ameloblastin—that stimulate dental follicle cells to differentiate into cementoblasts and form acellular extrinsic fiber cementum.^{8,15-17}

Pioneering experimental work by Hammarström and colleagues demonstrated that enamel matrix proteins applied to denuded root surfaces stimulated the regeneration of cementum, inserting Sharpey's fibers, and alveolar bone.^{11,18} These findings provided the first evidence that enamel matrix components could reproduce the biological environment of root

development, establishing the foundation for biomimetic periodontal regeneration.

Building upon these principles, a commercially usable formulation—Emdogain®—was developed in the 1990s. This preparation contains approximately 90% porcine amelogenin proteins suspended in a propylene glycol alginate carrier, designed to mimic the natural enamel matrix secreted during early root development. Subsequent clinical introduction and regulatory approval marked a major advancement in biologically driven periodontal therapy, enabling clinicians to harness natural cementogenesis pathways to enhance regenerative outcomes.^{6,19,20}

Over the past three decades, EMD has become one of the most extensively studied biologic agents in periodontology, with consistent evidence supporting its role in promoting acellular cementum formation, PDL fiber insertion, and alveolar bone regeneration.

Biochemical Composition of Enamel Matrix Derivative (EMD)

Enamel matrix derivative (EMD) is a biologically active extract obtained from developing porcine tooth buds and is composed primarily of proteins involved in enamel and cementum formation. Its major constituent is amelogenin, which accounts for nearly 90% of the total protein content. Amelogenins play a key regulatory role in enamel biomineralization and early cementogenesis by influencing cell adhesion, proliferation, and differentiation.^{6,17}

In addition to amelogenin, EMD contains several minor enamel matrix proteins, including ameloblastin, enamelin, tuftelin-related peptides, and other biologically active components that contribute to extracellular matrix organization and mineral deposition.^{7,8} These proteins collectively mimic the native enamel matrix secreted by Hertwig's epithelial

root sheath (HERS) during root development, thereby reproducing the biological environment necessary for cementoblast differentiation and acellular cementum formation.

The protein mixture is suspended in a propylene glycol alginate (PGA) carrier solution, which enhances clinical handling characteristics and facilitates sustained release and stability on conditioned root surfaces. This formulation allows EMD to be applied effectively during periodontal regenerative procedures, maintaining a biologically conducive environment for cell–matrix interactions.²⁰

The biomimetic composition of EMD underlies its ability to induce cementogenesis, support PDL regeneration, and promote alveolar bone formation, aligning closely with the natural sequence of periodontal tissue development.

Biological and Molecular Mechanisms of EMD

Enamel matrix derivative (EMD) exerts its regenerative effects through a combination of cellular responses and activation of key molecular signaling pathways. These mechanisms collectively promote cementogenesis, periodontal ligament (PDL) regeneration, and alveolar bone formation, closely mimicking natural root development.

Cellular Effects

EMD influences several types of periodontal cells involved in regeneration:

➤ Periodontal Ligament Fibroblasts

EMD enhances PDL fibroblast adhesion, spreading, migration, and proliferation, facilitating early wound stabilization and extracellular matrix formation.^{7,9} These cells synthesize collagen type I and III, contributing to functional PDL fiber insertion.

➤ Cementoblasts

Cementoblasts exposed to EMD exhibit increased alkaline phosphatase activity, mineralized nodule formation, and cementum-related gene expression, promoting acellular extrinsic fiber cementum formation.^{17,21}

➤ Osteoblasts and Bone-Forming Cells

EMD stimulates osteoblast differentiation, collagen matrix production, and expression of bone sialoprotein and osteocalcin, contributing to alveolar bone regeneration adjacent to treated root surfaces.^{7,22}

➤ Stem Cells of the Periodontal Ligament (PDLSCs)

PDLSCs demonstrate enhanced proliferation and cementogenic/osteogenic differentiation when exposed to amelogenin peptides, reflecting the biomimetic effect of enamel matrix components on multipotent precursor cells.²³

➤ Angiogenesis and Inflammatory Modulation

EMD upregulates vascular endothelial growth factor (VEGF), supporting angiogenesis essential for early healing. Simultaneously, it reduces pro-inflammatory mediators such as IL-1 β and TNF- α , shifting the wound environment toward regeneration rather than repair.^{9,24}

Molecular Pathways Activated by EMD

Multiple signaling pathways are activated by EMD, contributing to periodontal tissue regeneration:

• TGF- β Signaling Pathway

EMD enhances TGF- β 1 expression, resulting in increased collagen production, extracellular matrix maturation, and fibroblast differentiation. This pathway is essential for early wound stabilization and cementoblast function.^{7,9}

• BMP-2/7 Pathways

Bone morphogenetic proteins BMP-2 and BMP-7 are upregulated following EMD application, promoting cementogenic and osteogenic differentiation by

activating Runx2 and other key transcription factors involved in mineralized tissue development.^{21,22}

- MAPK Pathway

Mitogen-activated protein kinase (MAPK) signaling, particularly ERK1/2, is stimulated by EMD, enhancing cell proliferation, cytoskeletal organization, and mineralized tissue formation.^{7,9}

- Wnt/ β -Catenin Signaling

EMD modulates canonical Wnt signaling, contributing to cementoblast differentiation and periodontal matrix organization. Wnt-associated pathways are strongly implicated in the development of periodontal tissues and regeneration.^{17,25}

Extracellular Matrix Synthesis and Remodeling

EMD increases expression of collagen type I, fibronectin, and non-collagenous proteins involved in cementogenesis. It also promotes matrix organization by influencing integrin expression and cytoskeletal structure.⁷

Histologic and Preclinical Evidence

Extensive histologic and preclinical research has demonstrated the regenerative potential of enamel matrix derivative (EMD) in periodontal tissues. These studies consistently confirm its ability to induce cementogenesis, stimulate periodontal ligament (PDL) fiber insertion, and promote alveolar bone formation—hallmarks of true periodontal regeneration.

Evidence from Animal Studies

- Cementum Formation

In canine, primate, and rodent models, EMD application on denuded root surfaces results in extensive formation of acellular extrinsic fiber cementum, considered the gold standard indicator of periodontal regeneration. This cementum exhibits the characteristic insertion of Sharpey's fibers, indicating functional attachment of PDL fibers.^{11,26,27}

- Periodontal Ligament Regeneration

Histologic sections reveal well-organized PDL fibers oriented perpendicular to the root surface, resembling native periodontal ligament architecture. EMD-treated sites consistently demonstrate improved collagen fiber orientation compared with non-treated controls.²⁶

- Alveolar Bone Formation

EMD stimulates significant alveolar bone regeneration adjacent to treated roots. Increased osteoblastic activity, bone matrix deposition, and mineralized tissue formation have been documented in multiple preclinical studies.²⁷

- Reduced Root Resorption and Ankylosis

An important advantage observed in preclinical models is the decrease in root resorption and ankylosis, compared with repair-dominated healing in untreated sites. EMD appears to stabilize the root surface and promote biological processes conducive to cementogenesis, reducing adverse healing events.²⁸

Histologic Evidence in Human Studies

- ✓ Intrabony Defects

Human histologic reports demonstrate formation of new cementum, PDL, and alveolar bone following EMD application in intrabony defects. These findings confirm that the regenerative effects observed in animal models translate effectively to human tissues.^{12,13}

- ✓ Furcation Defects

EMD-treated furcation defects exhibit connective tissue attachment and new bone formation, although outcomes vary depending on defect morphology and surgical technique.¹²

- ✓ Soft-Tissue Regeneration

Histologic sections from recession coverage procedures show new cementum formation and insertion of collagen fibers, indicating biologically driven root coverage rather than epithelial adaptation.¹³

Factors Influencing Histologic Outcomes

- Several variables influence the extent of histologic regeneration:
- Defect morphology (narrow and deep defects respond best)
- Wound stability during early healing
- Inflammatory status of surrounding tissues
- Host healing capacity
- Surgical technique and operator precision
- Environmental sensitivity of amelogenins (pH, temperature, contamination)

Although EMD demonstrates strong regenerative potential, these factors contribute to variat

Combined and Synergistic Applications of EMD

Enamel matrix derivative (EMD) has demonstrated significant potential when used alone; however, numerous studies indicate that its regenerative capacity can be further enhanced when combined with adjunctive biomaterials, growth factors, and scaffolds. These synergistic approaches aim to improve defect fill, wound stability, space maintenance, and cellular recruitment—factors crucial for achieving predictably successful regeneration.

EMD Combined with Bone Grafts

The combination of EMD with xenografts or allografts aims to overcome limitations of space collapse and insufficient defect stability. While EMD promotes cementogenesis and PDL fiber insertion, bone grafts provide a three-dimensional scaffold that enhances bone volume and defect fill.

Clinical and preclinical studies report improved radiographic bone fill and attachment levels when EMD is used with deproteinized bovine bone mineral (DBBM) compared with EMD alone.^{4,12}

EMD with Collagen Membranes or Matrices

Collagen membranes offer additional wound stability and compartmentalization. When used with EMD, they support early healing through matrix-guided tissue formation.

Collagen matrices also improve soft-tissue volume in root-coverage procedures by serving as a scaffold for cellular infiltration and angiogenesis.^{14,29}

EMD Combined with Growth Factors

Platelet-Derived Growth Factor (PDGF)

PDGF promotes chemotaxis and proliferation of periodontal fibroblasts. When used with EMD, it enhances extracellular matrix formation and early wound stabilization. Both biomolecules act through complementary pathways—EMD via amelogenin-derived signals and PDGF via receptor-mediated mitogenesis—resulting in improved regenerative outcomes.

Bone Morphogenetic Proteins (BMP-2, BMP-7)

Since EMD upregulates BMP activity intrinsically,^{21,22} combining it with exogenous BMPs may amplify cementogenic and osteogenic differentiation. Preclinical models suggest increased mineralized tissue deposition, although clinical evidence remains limited.

EMD with Tissue Engineering and Biomimetic Scaffolds

Advanced biomaterials such as nanofibrous scaffolds, hydrogels, and 3D-printed constructs have been explored as carriers for amelogenin peptides. These platforms improve protein stability, prolong release, and direct cell migration and differentiation.

Hydrogel-based carriers help overcome EMD's sensitivity to pH and temperature fluctuations, while nanostructured scaffolds provide enhanced cell adhesion and growth factor incorporation.^{9,24}

Recombinant and Synthetic Amelogenin-Based Systems

Recombinant amelogenin peptides and synthetic enamel matrix protein analogs attempt to isolate the active domains of EMD responsible for cementogenesis. These agents may eliminate:

- variability associated with porcine-derived proteins
- immunogenicity concerns
- storage and environmental limitations
- Early in vitro results show comparable cementogenic and osteogenic effects with improved biochemical stability.^{21,23}

Clinical Implications of Synergistic Approaches

The combination of EMD with adjunctive materials provides several advantages:

- Improved space maintenance for bone regeneration
- enhanced soft-tissue volume and stability
- Improved biological signaling through multi-pathway activation
- Better defect fill and long-term attachment gain
- Increased predictability in challenging defects (wide, shallow, or combined defects)

However, clinical outcomes vary depending on defect morphology, biomaterial selection, and operator technique.

Limitations of Enamel Matrix Derivative (EMD)

Although enamel matrix derivative (EMD) is one of the most widely used biologics in periodontal regeneration, several limitations influence its predictability, handling characteristics, and overall clinical performance. Understanding these limitations is essential for appropriate case selection and optimizing treatment outcomes.

Sensitivity to Environmental Conditions

Amelogenins, the principal components of EMD, are highly sensitive to environmental changes:

pH variations can destabilize protein structure and reduce bioactivity.

Temperature fluctuations may alter viscosity and protein conformation.

Contamination with blood or saliva can interfere with protein adsorption onto the root surface.^{7,9}

Because amelogenins require a stable environment, improper handling may compromise regenerative potential.

Variable Clinical Outcomes

Although many studies show favorable results, clinical performance is not uniformly predictable:

- Wide, shallow, or non-contained defects respond less favorably than narrow and deep defects.
- Furcation defects show inconsistent outcomes due to anatomical complexity.^{10,12}
- Differences in surgical skill and wound stabilization significantly influence regenerative success.

This variability limits the universal applicability of EMD in all defect types.

Limited Space-Making Ability

EMD is a gel-based biomaterial, which means it: does not provide structural support, cannot maintain space in large or unsupported defects, may collapse without additional scaffolding (e.g., xenografts or membranes).^{4,12}

Therefore, adjunctive materials are often required for predictable bone regeneration.

Porcine-Derived Origin

Because EMD is derived from porcine tooth buds, ethical, cultural, and religious concerns may limit its acceptability for certain patients or populations. Additionally:

Some patients may prefer synthetic or recombinant alternatives.

Although rare, concerns about immunogenicity exist, even though extensive clinical data show it is safe. [6,20]

The porcine origin remains a practical and ethical limitation.

Cost Considerations

EMD is relatively expensive compared with other regenerative materials. Cost affects:

patient acceptance,

feasibility in large defects where greater material volume is required,

economic justification for combining EMD with additional grafts or membranes.

In low-resource clinical settings, its routine use may be restricted.

Lack of Strong Evidence for Some Combinations

Although synergistic benefits have been suggested, robust controlled trials are still lacking for:

EMD with BMPs, PDGF, or multiple biologics

EMD with emerging biomimetic scaffolds

EMD in large-scale tissue-engineered constructs [9,24]

More high-quality randomized controlled trials are needed to validate advanced combination therapies.

Limited Longevity of Evidence in Certain Applications

While EMD shows consistent results in intrabony defects and recession coverage, evidence is less conclusive for:

Class III furcation defects

Complex combined defects

Peri-implant defects (due to limited biologic rationale) [13,14]

This restricts EMD's role to defect types with established efficacy.

Dependence on Root Surface Conditioning

Optimal EMD function requires conditioned root surfaces, typically treated with EDTA:

Removes smear layer

Exposes collagen fibrils

Enhances protein adsorption⁷

However, improper conditioning (over- or under-conditioning) can impair protein binding and limit regeneration.

Future Directions and Emerging Innovations

Despite the well-established role of enamel matrix derivative (EMD) in periodontal regeneration, ongoing advancements in biomaterials science, protein engineering, and tissue regeneration offer promising opportunities to further enhance its clinical performance. Future research is increasingly focused on optimizing delivery systems, improving protein stability, and developing biologically superior alternatives or adjuncts to conventional EMD.

Recombinant and Synthetic Amelogenin-Based Biomolecules

The development of recombinant amelogenin peptides represents a major step forward in overcoming:

porcine-origin limitations,

variability in natural protein mixtures,

storage instability under environmental fluctuations.

These synthetic peptides reproduce the functional domains responsible for cementogenesis and PDL regeneration, offering greater purity, consistency, and biocompatibility. [21,23]

Advanced Delivery Systems: Hydrogels, Nanocarriers, and Controlled Release Platforms

Current research is directed toward next-generation carriers that improve EMD stability and bioavailability.

These include:

thermo-responsive hydrogels,

nanofiber scaffolds,

microsphere-based slow-release systems,

biofunctionalized 3D-printed scaffolds.

These innovations help maintain optimal protein activity, prolong release, enhance cellular infiltration, and provide structural support for defect stability. [9,24]

Combination Therapies with Next-Generation Growth Factors

Emerging biologics—such as FGF-2, IGF-1, and SDF-1—offer novel synergistic potential with amelogenin-based molecules. Rational combination therapy may target multiple phases of periodontal wound healing:

chemotaxis (SDF-1),
angiogenesis (FGF-2),
osteogenesis (BMPs),
extracellular matrix maturation (EMD).

Synergistic multi-factor systems may create a more robust regenerative microenvironment.

Tissue Engineering and Cell-Based Regeneration

EMD's ability to stimulate periodontal ligament stem cells (PDLSCs) opens opportunities for:

cell-sheet engineering,
scaffold-free cementoblast constructs,
stem-cell biomolecule hybrid therapies.

Integrating EMD with stem-cell-based methods may enhance regeneration in complex defects not responsive to conventional therapies.

Gene-Activated and Bio-Instructive Scaffolds

Future platforms may incorporate:
gene delivery vectors encoding amelogenin or BMPs,
smart scaffolds that release signals in response to pH or inflammatory cues,
bio-instructive surfaces that guide cementoblast and fibroblast orientation.

These technologies aim to reproduce the spatial and temporal dynamics of natural periodontal development.

Personalized and Precision Regeneration

With emerging tools such as:
salivary proteomics,

microbiome analysis,
genetic risk profiling,

future periodontal regeneration may be tailored to individual wound biology, optimizing EMD selection, dosage, and adjunctive materials for personalized therapy.

Conclusion

Enamel matrix derivative (EMD) remains one of the most extensively validated biologic agents for periodontal regeneration. Its ability to stimulate acellular extrinsic fiber cementum, promote periodontal ligament fiber insertion, and enhance alveolar bone formation reflects its biomimetic alignment with natural root development. EMD exerts multifaceted effects, including cellular proliferation, differentiation, extracellular matrix synthesis, angiogenesis, and modulation of inflammatory responses—mechanisms that collectively support regenerative rather than reparative healing.

Histologic, preclinical, and clinical evidence demonstrates consistent success in intrabony defects, recession coverage procedures, and selected furcation involvements. However, limitations such as environmental sensitivity, variable clinical outcomes, lack of intrinsic space maintenance, and porcine origin highlight the need for careful case selection and appropriate combination with adjunctive biomaterials.

Emerging strategies—including recombinant amelogenin peptides, nanostructured carriers, engineered scaffolds, and multi-factor regenerative systems—offer exciting avenues to enhance the predictability and biological performance of EMD. As periodontal regenerative science advances, EMD will continue to serve as a foundational biologic while inspiring new generations of biomimetic and bioengineered therapies.

In summary, EMD represents a safe, biologically potent, and clinically versatile regenerative material, with future innovations poised to further refine its role in modern periodontal therapy.

References

1. Melcher AH. On the repair potential of periodontal tissues. *J Periodontol.* 1976;47(5):256–260.
2. Nyman S, Gottlow J, Karring T, Lindhe J. The regenerative potential of the periodontal ligament. *J Clin Periodontol.* 1982;9(3):257–265.
3. Caton J, Greenstein G. Factors related to periodontal regeneration. *Periodontol 2000.* 1993;1:9–15.
4. Sculean A, Nikolidakis D, Schwarz F. Regeneration of periodontal tissues using regenerative materials. *Clin Oral Investig.* 2015;19(7):1433–1446.
5. Hammarström L. Enamel matrix and cementum development. *J Clin Periodontol.* 1997;24(9 Pt 2):658–668.
6. Gestrelus S, Andersson C, Johansson AC, et al. Formulation of enamel matrix derivative. *J Clin Periodontol.* 1997;24(9):678–684.
7. Bosshardt DD. Enamel matrix proteins: cellular and molecular influence in periodontal regeneration. *J Clin Periodontol.* 2008;35(Suppl 8):87–105.
8. Fincham AG, Moradian-Oldak J, Simmer JP. The structural biology of enamel matrix proteins. *J Struct Biol.* 1999;126(3):270–299.
9. Grandin HM, Gemperli AC, Dard M. Enamel matrix derivative and its effects on periodontal wound healing. *Tissue Eng Part B Rev.* 2012;18(3):181–202.
10. Heijl L, Heden G, Svardström G, Ostgren A. Enamel matrix derivative in the treatment of intrabony periodontal defects. *J Clin Periodontol.* 1997;24(9):705–714.
11. Hammarström L, Heijl L, Gestrelus S. Periodontal regeneration in monkeys following EMD application. *J Clin Periodontol.* 1997;24(9):669–677.
12. Camelo M, Nevins ML, Schenk RK, Lynch SE, Nevins M. Periodontal regeneration with enamel matrix derivative. *Int J Periodontics Restorative Dent.* 2003;23(2):157–165.
13. Sculean A, Donos N, Windisch P, et al. Healing of intrabony defects with enamel matrix proteins or guided tissue regeneration. *J Periodont Res.* 1999;34(8):310–322.
14. Sculean A, Gruber R, Bosshardt DD. Various roles of EMD in periodontal wound healing. *J Clin Periodontol.* 2015;42(Suppl 16):S187–202.
15. Termine JD, Belcourt AB, Christner PJ, Conn KM, Nysten MU. Fetal tooth matrix proteins and cementogenesis. *Calcif Tissue Res.* 1980;31(1):45–56.
16. Nanci A, Bosshardt DD. Structure of periodontal tissues. *Periodontol 2000.* 2006;40:11–28.
17. Zeichner-David M et al. Hertwig's epithelial root sheath and root development. *Dev Dyn.* 2003;228:651–660.
18. Slavkin HC, Boyde A. Cementum as a possible epithelial product. *J Dent Res.* 1975;54(Spec No):138–153.
19. Heijl L et al. Enamel matrix derivative in periodontal defects. *J Clin Periodontol.* 1997;24:705–714.
20. Lyngstadaas SP, Risnes S, Sproat BS et al. Bioactivity of enamel matrix proteins. *Clin Oral Investig.* 2000;4:120–125.
21. Diekwisch TG. Developmental biology of cementum. *Int J Dev Biol.* 2001;45:695–706.

22. Hoang AM, Oates TW, Cochran DL. Amelogenin enhances osteoblast-related gene expression. *J Periodontol.* 2002;73:419–425.
23. Lyngstadaas SP, Risnes S, Sproat BS et al. Amelogenin peptides cause dose-dependent differentiation of periodontal ligament cells. *J Dent Res.* 2001;80:2082–2087.
24. Grandin HM, Gemperli AC, Dard M. Cellular and molecular effects of enamel matrix derivative. *Tissue Eng Part B Rev.* 2012;18:181–202.
25. Sonoyama W, Seo BM, Yamaza T, Shi S. Hertwig's epithelial root sheath cells play key roles in cementum formation. *J Dent Res.* 2007;86:594–599.
26. Bosshardt DD, Nanci A. Cementogenesis revisited. *J Periodont Res.* 2004;39:348–356.
27. Donos N, Sculean A, Glavind L, Reich E, Karring T. Regenerative cementum formation after EMD application. *J Clin Periodontol.* 2006;33:437–443.
28. Heden G, Wennström JL, Lindhe J. Periodontal tissue alterations following application of enamel matrix derivative. *J Clin Periodontol.* 2001;28:342–349.
29. McGuire MK, Scheyer ET. A randomized controlled clinical trial evaluating EMD with a collagen matrix in soft-tissue augmentation. *J Periodontol.* 2016;87:435–444.